

(12) INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

(19) World Intellectual Property Organization  
International Bureau



(43) International Publication Date  
14 August 2003 (14.08.2003)

PCT

(10) International Publication Number  
WO 03/066682 A1

(51) International Patent Classification<sup>7</sup>: C08B 37/08

(21) International Application Number: PCT/IB03/00074

(22) International Filing Date: 9 January 2003 (09.01.2003)

(25) Filing Language: English

(26) Publication Language: English

(30) Priority Data:  
P.352079 7 February 2002 (07.02.2002) PL

(71) Applicant (for all designated States except US): **ABBOTT LABORATORIES DE COSTA RICA LTD** [BS/BS];  
Sassoon House, Shirley Street and Victoria Avenue,  
Nassau, Island of New Providence (BS).

(72) Inventors; and

(75) Inventors/Applicants (for US only): **STRUSZCZYK, Henryk** [PL/PL]; ul. Tuwima 8 m.29, 95-100 Zgierz (PL). **KUCHARSKA, Magdalena** [PL/PL]; ul. Kostki Napier-  
skiego 2 m.39, 94-056 Lodz (PL). **NIEKRASZEWICZ, Antoni** [PL/PL]; ul. Wici 72 m.9, 91-157 Lodz (PL). **UR-  
BANOWSKI, Alojzy** [PL/PL]; ul. Limbowa 41, 92-015  
Lodz (PL). **WESOŁOWSKA, Ewa** [PL/PL]; ul. Elsnera  
9 m.36, 92-504 Lodz (PL). **CIECHANska, Danuta**  
[PL/PL]; ul. Bialostocka 25 m.1, 93-355 Lodz (PL).

(81) Designated States (*national*): AE, AG, AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, BZ, CA, CH, CN, CO, CR, CU, CZ, DE, DK, DM, DZ, EC, EE, ES, FI, GB, GD, GE, GH, GM, HR, HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MA, MD, MG, MK, MN, MW, MX, MZ, NO, NZ, OM, PH, PL, PT, RO, RU, SD, SE, SG, SK, SL, TJ, TM, TN, TR, TT, TZ, UA, UG, US, UZ, VC, VN, YU, ZA, ZM, ZW.

(84) Designated States (*regional*): ARIPO patent (GH, GM, KE, LS, MW, MZ, SD, SL, SZ, TZ, UG, ZM, ZW), Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE, BG, CH, CY, CZ, DE, DK, EE, ES, FI, FR, GB, GR, HU, IE, IT, LU, MC, NL, PT, SE, SI, SK, TR), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, GQ, GW, ML, MR, NE, SN, TD, TG).

Declaration under Rule 4.17:

— of inventorship (Rule 4.17(iv)) for US only

Published:

— with international search report

For two-letter codes and other abbreviations, refer to the "Guidance Notes on Codes and Abbreviations" appearing at the beginning of each regular issue of the PCT Gazette.

(54) Title: METHOD FOR DEPROTEINIZATION OF CHITOSAN

(57) Abstract: Methods for deproteinizing chitosan are provided. Proteins are dissolved in aqueous solution under intensive agitation. Precipitated agglomerated microcrystalline product is subject to aqueous acidic or basic salt solution, from which dissolved proteins can be removed. Residual product is deproteinized chitosan.

WO 03/066682 A1

5

## METHOD FOR DEPROTEINIZATION OF CHITOSAN

### FIELD OF THE INVENTION

10

The invention concerns a method for deproteinization of chitosan.

### BACKGROUND OF THE INVENTION

15

20

25

30

35

40

45

50

"Journal of Applied Polymer Science", vol. 2, p. 380, 1980; "Fishery Technology", vol. 11, p.50, 1974; "Carbohydrate Research", vol. 38, p.35, 1974, "Journal of Food Science Technology", vol. 12, p.187, 1975; "Journal of Organic Chemistry", vol. 27, p. 161, 1962; "Biotechnology & Bioengineering", vol. 20, p. 1931, 1978; "Journal of Agriculture and Food Chemistry", vol. 37, p. 5 - 75, 1989; "Journal of Agriculture and Food Chemistry", vol. 39, p. 1527, 1991; "Acta Polymer", vol. 45, p. 41, 1994; "Food Biotechnology", vol. 7, p. 253, 1993 and U.S. Patents 3,533,940, 3,862,122, 3,922,260, 4,066,735, 4,195,175, and 4,199,496 and Polish Patent 119,931 and 160,714 teach methods to reduce the protein content in chitin and chitosan. These methods are based on the treatment of shells of crustaceans and insects with aqueous alkali such as sodium hydroxide, potassium hydroxide, calcium hydroxide, or their salts as sodium carbonate, sodium hydrogen carbonate, sodium sulfite, sodium hydrogen sulfite, sodium sulfide or sodium phosphate, with concentration of 0.5 - 10.0 wt % at temperature of 20 - 150°C for 1 - 72 h. These well - known methods allow the reduction of the protein content in chitin and chitosan, however, they are not able to eliminate proteins from chitin and chitosan.

U.S. Patents 5,623,064 and 5,624,679 teach methods to produce chitin and chitosan with high purity without proteins. These methods are based on mechanical or enzymatic treatment of microalgae strain of *Coscinodiscus* genus, *Cyclotella* genus and *Thalassiosira* genus. Microalgae biomass is treated by aqueous solution of hydrochloric acid at temperature 70 °C, water and ethyl alcohol whereas the proteins are removed by treatment with surfactants such as sodium dodecylsulphonate. This well - known method is characterized by low yield and the tendency of polyaminosaccharides toward hydrolytic degradation in acidic medium. This method is not useful for deproteinization of chitin and chitosan originated from sources like the shells of crustaceans and insects.

U.S. Patent 5,053,113 teaches electrochemical deproteinization and demineralization of raw materials containing chitin. These chitin based raw materials are treated with 0.1 - 2.0 % aqueous sodium hydroxide using electric current of 4 - 11 A and voltage of 15 - 50V for 10 - 45 min.

U.S. Patent 6,310,188 teaches a method to produce chitin and chitosan by transformation of crustacean shells into amorphous form. The crustacean shells are heated at 78°C and drastically cooled in liquid nitrogen. The amorphous shells are used to prepare chitin, and may be deproteinized with aqueous sodium hydroxide.

These well - known methods do not allow production of chitin with protein content lower than 10 - 20 ppm.

## 5 SUMMARY OF THE INVENTION

Disclosed are methods for deproteinizing chitosan by precipitating microcrystalline chitosan from aqueous solution of chitosan.

10

## DESCRIPTION OF THE INVENTION

The method for deproteinization of chitosan according to the invention, consists in that the chitosan, containing proteins dissolved in aqueous solutions of acids such as hydrochloric, acetic or lactic, with polymer concentration not lower than 0.001 wt%, preferably 0.5 - 2.0 wt%, is agglomerated using aqueous solutions of base or / and its salts for not less than 1 minute, preferable 30 - 120 minutes, with intensive agitation at 100 - 1000 rpm. Precipitated agglomerated microcrystalline product is subjected to aqueous base or basic salt solution with concentration not lower than 0.1 wt%, preferably 1 - 10 wt%, for time ranged from 1 minute to 100 hours. Then, the aqueous base or basic salt solution containing dissolved proteins is removed from reaction medium and the residual product is washed by water and / or alcohol, preferably ethyl alcohol, to remove all contaminants and the resulted chitosan is concentrated and possibly dried by well - known methods.

25

The chitosan agglomeration process according to the invention can be carried out in two steps. The first step involves addition of aqueous solution of base or / and its salts to obtain a reaction medium pH = 6.0 --6.5. Then a second aqueous solution of base or/and its salts is added. The ratio of concentration of alkali in the first basic solution relative to that in the second alkali solution is 1: 0.1 to 1 : 0.9.

30

Aqueous solutions of sodium or potassium hydroxide or/ and their salts such as sodium or potassium carbonate are used in the method according to the invention. The aqueous solution of base or / and its salts containing dissolved proteins in the method according to invention is removed from reaction medium by filtration, ultrafiltration, sedimentation or centrifugation.

35

Deproteinization of chitosan in accordance to the invention is achieved by removing proteins from agglomerated microcrystalline chitosan by their dissolution in aqueous alkali. Specific structure of agglomerated chitosan, including its porosity, water retention value higher than 500 % as well as developed intrinsic surface, microcapillary and capillary system, support alkali diffusion into chitosan structure, resulting in protein dissolution. A method according to the invention destroys the stable complex connections of proteins with initial chitosan by its dissolution followed by agglomeration. A two stage agglomeration allows production of the agglomerated chitosan with special developed intrinsic surface accessible for alkali penetration and protein dissolution.

40

45

A benefit of the method according to the invention is to maximize removal of proteins from developed structure of chitosan by treatment with aqueous bases or/and their salts as well as by washing using water with reduced reaction medium of agglomerated chitosan. This process supports removal of the protein from chitosan structure.

50

5       The structure of resultant microcrystalline chitosan is specially susceptible on solvent exchange processes, including alkali treatment acting positively for protein removal.

10       Chitosan deproteinized according to the invention is characterized by high degree of purity and protein content lower than 10 ppm. This chitosan is widely applied in medicine, pharmacy or biotechnology.

15       The method according to the invention is illustrated with following examples, which do not limit its range of application.

#### EXAMPLE I

20       99 weight parts of 0.4 wt% aqueous hydrochloric acid solution and 1 weight part of chitosan flakes with viscometric average molecular weight  $M_v = 796$  kD, deacetylation degree  $DD = 85.6\%$ , water retention value  $76.6\%$ , moisture content  $11.1\%$ , ash content  $0.21\%$  and protein content of  $350$  ppm were introduced into reactor equipped with agitator and cooling jacket. Chitosan was dissolved for  $2$  h with agitation of  $120$  rpm, then the solution was filtered on the frame filter, obtaining  $96$  weight parts of  
25       solution containing  $0.99$  wt % of chitosan. This solution was transferred into a reactor equipped with high speed agitator and cooling jacket where this solution with agitation of  $480$  rpm was treated with gradual addition of  $53.5$  weight parts of  $0.75\%$  aqueous solution of sodium hydroxide to obtain a reaction medium  $pH = 8.20$  and precipitation of agglomerated microcrystalline product in a form of dispersion. The resulting dispersion was  
30       concentrated on the nutsche filter to obtain  $40$  weight parts of agglomerates that were transferred to a previous reactor containing  $80$  weight parts of  $5.0\%$  aqueous sodium hydroxide. A process of protein removal was carried out for  $3$  h at temperature  $20^\circ C$  with agitation of  $30$  rpm. The dispersion was next concentrated on the nutsche filter obtaining  $40$  weight parts of chitosan dispersion. This dispersion was transferred again to the  
35       reactor containing  $40$  weight parts of demineralized water with  $pH = 6.50$ . A content of reactor was homogenized for  $15$  minutes and filtered with estimation of protein concentration in filtrate and chitosan dispersion. A chitosan dispersion was washed  $20$  times on the nutsche filter using demineralized water to eliminate the presence of protein in filtrate and to obtain the reaction medium  $pH = 7.35$  in the microcrystalline  
40       agglomerate.

$30$  weight parts of protein - free chitosan agglomerates containing  $3.15$  wt% of polymer with  $M_v = 750$  kD,  $DD = 680\%$  and ash content  $0.12\%$  were obtained.

#### EXAMPLE II

50        $99$  weight parts of  $2.0\%$  aqueous solution of acetic acid and  $1.2$  weight parts of chitosan flakes with properties as in Example I were introduced in the reactor as in Example I. Chitosan was dissolved for  $3$  h with agitation rate of  $120$  rpm, then the chitosan solution was filtered on the frame filter to obtain  $96$  weight part of solution containing  $1.15\%$  of chitosan. This solution was transferred into reactor with high speed agitator and cooling jacket and  $88$  weight parts  $1.5$  aqueous sodium hydroxide solution was gradually added to the reactor with continuous agitation with  $500$  rpm to obtain a reaction medium  $pH = 8.20$ .

5 Agglomerated microcrystalline product in a form of dispersion. This dispersion was concentrated on the nutsche filter precipitated to obtain 40 weight parts of agglomerates that were transferred to a reactor equipped with low speed agitator and 80 weight parts 5.0% aqueous sodium hydroxide. Treatment of chitosan agglomerate was carried out for 3 h at temperature 20°C with agitation rate 30 rpm. Then, the mixture was concentrated on the  
10 nutsche filter obtaining 40 weight parts of chitosan dispersion. This dispersion was mixed for 15 minutes with 40 weight parts of demineralized water with pH = 6.50 and next a content of reactor was filtered with estimation of protein concentration in filtrate and chitosan agglomerate. The washing process by demineralized water was repeated on the nutsche filter 20 times to eliminate the protein in filtrate and obtain the  
15 chitosan agglomerate reaction pH = 7.40.

28 weight parts of protein - free chitosan agglomerates were obtained with polymer content 3.41 wt%,  $M_v = 760$  kD, DD = 85.6 %, WRV = 820 % and ash content 0.12 %.

20

### EXAMPLE III

99 weight parts of aqueous lactic acid solution and 1 weight part of chitosan powder with  $M_v = 400$  kD, DD = 79.2 %, moisture content 5.68 %, ash content 1.3 %, WRV = 244 % and protein content of 600 ppm was agitated with 100 rpm for 2.5 h in the reactor as in Example I. This solution was filtered on the frame filter to obtain 95 weight parts of chitosan solution containing 0.98 % of polymer. This filtered solution was transferred to a reactor equipped with high speed agitator and cooling jacket and 53.56  
30 weight parts of 0.5 % aqueous potassium hydroxide solution was added gradually with agitation with 400 rpm to obtain a reaction medium pH = 8.00 and precipitation of agglomerated product in a microcrystalline form. This dispersion was concentrated using filtration centrifuge to obtain 230 weight parts of chitosan agglomerate, which was transferred to a reactor equipped with agitator containing 60 weight parts 5.0 % aqueous potassium hydroxide solution then a deproteinization was carried out for 5 h with agitation rate of 30 rpm at temperature 20°C. The resulting dispersion was concentrated using  
35 centrifuge to obtain 30 weight parts of chitosan agglomerate dispersion. This chitosan dispersion was transferred again to a reactor containing 30 weight parts of demineralized water with pH = 6.50. The mixture was agitated for 15 minutes and next filtered with estimation of protein concentration in filtrate and chitosan agglomerate. The washing process by demineralized water was repeated on the filtering antifuge 22 times to obtain the  
40 chitosan agglomerate reaction of pH = 7.30.

33 weight parts of chitosan agglomerates with polymer content 2.92 wt%,  $M_v = 370$  kD, DD = 79.2 %, ash content 0.22 %, WRV = 1050 % and protein content lower than 10 ppm were obtained. This product is acceptable for medical and pharmaceutical uses.

### EXAMPLE IV

50 99 weight parts of 0.4 % aqueous hydrochloric acid solution and 1 weight part of chitosan flakes with  $M_v = 850$  kD, DD = 83.4 %, moisture content 13.0%, ash content 0.59%, WRV = 139 % and protein content of 1750 ppm were agitated for 2 h with 280 rpm in the reactor as in Example I. The resulting solution was filtered on the frame filter to obtain 96 weight parts of chitosan solution containing 0.99 wt % of polymer. This chitosan

5 solution was transferred to the reactor equipped with high speed agitator and cooling jacket and 43.0 weight parts of 0.75 % aqueous sodium hydroxide was gradually added with agitation rate of 480 rpm to obtain a reaction medium pH = 6.46, then 16.0 weight parts of 0.50 % aqueous sodium hydroxide solution was gradually added with the same agitation rate to obtain a medium reaction pH = 7.99 and precipitation of agglomerated product in the microcrystalline form. The resulting dispersion was concentrated on the filtration nutsche to obtain 39 weight parts of chitosan agglomerates. These agglomerates were transferred to the reactor (equipped with an agitator) containing 78 weight parts 5.0 % aqueous sodium hydroxide; deproteinization was carried out for 5 h at temperature 20°C with agitation rate 30 rpm. This mixture was concentrated on the nutsche filter, transferred again to the reactor containing 40 weight parts of demineralized water with pH = 6.50 to agitate for 15 minutes. A mixture was filtered on the nutsche filter with estimation of protein content in filtrate and chitosan agglomerates. The washing process by demineralization was repeated 20 times to obtain a reaction medium pH = 7.35.

20 30 weight parts of chitosan agglomerates distinguished by protein content of 350 ppm were obtained in this stage. Deproteinization was repeated twice using 30 weight parts 1.0 % aqueous sodium hydroxide with agitation rate 30 rpm for 3 h at temperature 20°C. The resulting mixture was concentrated to obtain 35 weight parts of chitosan dispersion that was homogenized for 15 minutes with 35 weight parts of demineralized water with pH = 6.50. The washing process with demineralized water on the nutsche filter was repeated 12 times to obtain the agglomerated chitosan reaction of pH = 7.30.

30 The final product was 30 weight parts of chitosan agglomerates having a polymer content 3.15 %,  $M_v = 770$  kD, DD = 83.4 %, WRV = 900 %, ash content 0.15 % and protein content lower than 10 ppm. This product is acceptable for medical and pharmaceutical uses.

#### 35 EXAMPLE V

1.0 weight parts of chitosan flakes with  $M_v = 240$  kD, DD = 84.3 %, moisture content 6.76 %, ash content 1.1 %, WRV = 140 % and protein content 728 ppm and 49 weight parts of 0.4 % aqueous hydrochloric acid solution were introduced to the reactor as in Example I. Dissolution was carried out for 2.5 h with an agitation rate of 120 rpm, then resulting solution was filtered on the frame filter to obtain 48.5 weight parts of chitosan solution containing 1.92 % of polymer. This chitosan solution was transferred to the reactor, equipped with high speed agitator and cooling jacket, and 26.5 weight parts aqueous sodium hydroxide solution was gradually added with agitation rate 480 rpm to obtain a reaction medium pH = 8.20 and precipitation of chitosan agglomerates in a micro crystalline form. This dispersion was concentrated on the nutsche filter to obtain 35 weight parts of chitosan agglomerate dispersion that was transferred to the reactor containing 70 weight parts 5.0 % aqueous sodium hydroxide solution. The mixture obtained was agitated at 30 rpm for 3 h at 20°C, then the dispersion was concentrated on the nutsche filter to obtain 40 weight parts of chitosan dispersion. This dispersion was homogenized with 40 weight parts of demineralized water with pH = 6.50 for 15 minutes. The dispersion was then filtered on the nutsche filter with estimation of protein content in filtrate and chitosan dispersion. A washing process by demineralized water was repeated 20 times to eliminate proteins and obtain a chitosan dispersion reaction pH = 7.35.

5       The final product was 3.0 weight parts of chitosan agglomerates containing 3.20 % polymer characterized with  $M_v = 230$  kD,  $DD = 84.3$  %, ash content 0.25 %,  $WRV = 750$  % and protein content lower than 10 ppm were obtained. This product is acceptable for medical and pharmaceutical use.

#### 10       EXAMPLE VI

15       99 weight parts of 2.0 % aqueous solution of acetic acid and 1.2 weight parts of chitosan flakes with properties as in Example I was agitated for 3 h with 120 rpm in the reactor as in Example I. The dissolved chitosan was filtered on a frame filter to obtain 96 weight parts of chitosan solution containing 1.15 % of polymer. This solution was transferred to the reactor, equipped with high speed agitator and cooling jacket, and 88 weight parts of 1.5 % aqueous solution of sodium hydroxide and sodium carbonate, in a weight ratio of 2 : 1, was gradually added with agitation rate 500 rpm to 20 obtain a reaction medium of  $pH = 8.20$  and precipitation of chitosan agglomerates in a microcrystalline form. The resulting dispersion was concentrated by centrifugation to obtain 40 weight parts of chitosan agglomerates. This chitosan form was introduced into the reactor equipped with agitator containing 80 weight parts 7.5 % aqueous sodium hydroxide and deproteinization was carried out for 2 h at  $20^\circ C$  with agitation rate 30 25 rpm. The mixture was concentrated on the nutsche filter to obtain 40 weight parts of chitosan dispersion that was homogenized for 15 minutes with 80 weight parts of demineralized water with  $pH = 6.50$ . A mixture was then filtered with estimation of protein presence in filtrate and chitosan dispersion. A washing process by demineralized water was repeated 20 times to eliminate the protein presence in filtrate and to obtain the 30 chitosan agglomerate reaction  $pH = 7.40$ . The product was washed by 75 weight parts of ethyl alcohol to obtain  $pH = 7.20$ .

35       The final product was 28 weight parts of chitosan agglomerates with polymer content 3.41 %,  $M_v = 220$  kD,  $DD = 84.3$  %, ash content 0.15 %,  $WRV = 820$  % and protein content lower than 10 ppm were obtained. This product is acceptable for medical and pharmaceutical uses.

#### 40       EXAMPLE VII

45       1 weight part of chitosan flakes with properties as in Example I and 99 weight parts of 0.4 % aqueous hydrochloric acid solution were introduced into the reactor as in Example I. Dissolution was carried out for 2 h with agitation rate 120 rpm, then the solution was filtered on the frame filter to obtain 96 weight parts of chitosan solution containing 0.99 % polymer. This solution was transferred to the reactor, equipped with high speed agitator and cooling jacket, and 43 weight parts 0.75 % aqueous sodium hydroxide solution was gradually added with agitation rate 480 rpm to obtain a reaction medium of  $pH = 6.46$  and 21.7 weight parts of 0.50 % aqueous sodium hydroxide was added in the same conditions to obtain a medium reaction of  $pH = 8.03$  and 50 precipitation of chitosan agglomerates in a microcrystalline form. The dispersed chitosan agglomerate solution was concentrated on the nutsche filter to obtain 40 weight parts of product that was introduced into reactor, equipped with agitator, containing 80 weight parts 5.0 % aqueous sodium hydroxide solution. Deproteinization was carried out for 3 h at temperature  $20^\circ C$  with agitation rate of 30 rpm. This dispersion was concentrated on the

5 nutsche filter to obtain 40 weight parts of chitosan dispersion, which was homogenized  
for 15 minutes with 40 weight parts of demineralized water with pH = 6.50. The  
resulting mixture was filtered on the nutsche filter with estimation of protein presence in  
filtrate and chitosan dispersion. The washing process by demineralized water was  
10 repeated 20 times on the nutsche filter to eliminate a residual protein and to obtain a  
chitosan agglomerate reaction of pH = 7.35. This product was concentrated by  
centrifugation.

The final product was 30 weight parts of protein - free chitosan agglomerates with  
polymer content 3.15 %,  $M_v = 750$  1(D, DD = 85.6 %, WRV = 800 % and ash content 0.12  
15 were obtained.



5

## CLAIMS

1. A method for deproteinizing chitosan, comprising the steps of:
  - 10 a) reacting an acidic solution of chitosan, said chitosan containing proteins  $\geq 0.001$  wt%, with an aqueous base to precipitate microcrystalline chitosan; and
  - 15 b) separating said precipitated microcrystalline chitosan from dissolved proteins to produce a microcrystalline chitosan having a protein content  $\leq 10$  ppm.
2. A method according to claim 1, wherein said acidic solution of chitosan comprises an acid selected from the group consisting of hydrochloric acid, acetic acid and  
20 lactic acid.
3. A method according to claim 1, wherein said aqueous base is selected from the group consisting of sodium hydroxide, potassium hydroxide, sodium carbonate, and potassium carbonate.  
25
4. A method according to claim 1, wherein said reacting step is carried out at  $6.0 \leq \text{pH} \leq 6.5$ .
5. A method according to claim 1, wherein said reacting step further comprises adding a first aqueous basic solution to reach  $6.0 \leq \text{pH} \leq 6.5$  and then adding a  
30 second aqueous basic solution, wherein the concentration ratio of alkali in said first aqueous basic solution to said second aqueous basic solution is between 1:0.1 to 1:0.9.
6. A method according to claim 1, wherein said separating step is carried out using a method selected from the group consisting of filtration, ultrafiltration, sedimentation and centrifugation.  
35
7. A composition of matter, comprising a chitosan prepared according to a method  
40 according to claim 1.

# INTERNATIONAL SEARCH REPORT

Int. Application No  
PCT/IB 03/00074

## A. CLASSIFICATION OF SUBJECT MATTER

IPC 7 C08B37/08

According to International Patent Classification (IPC) or to both national classification and IPC

## B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

IPC 7 C08B

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the International search (name of data base and, where practical, search terms used)

WPI Data, PAJ, CHEM ABS Data, EPO-Internal

## C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	<p>PATENT ABSTRACTS OF JAPAN vol. 012, no. 223 (C-507), 24 June 1988 (1988-06-24) &amp; JP 63 017901 A (HIGETA SHOYU KK), 25 January 1988 (1988-01-25) abstract &amp; DATABASE WPI Section Ch, Week 198809 Derwent Publications Ltd., London, GB; Class D17, AN 1988-061314 &amp; JP 63 017901 A (HIGETA SHOKYU KK), 25 January 1988 (1988-01-25) abstract</p> <p style="text-align: center;">--- -/-</p>	1-4,6,7

☒ Further documents are listed in the continuation of box C.

☒ Patent family members are listed in annex.

### \* Special categories of cited documents :

- \*A\* document defining the general state of the art which is not considered to be of particular relevance
- \*E\* earlier document but published on or after the international filing date
- \*L\* document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)
- \*O\* document referring to an oral disclosure, use, exhibition or other means
- \*P\* document published prior to the international filing date but later than the priority date claimed

- \*T\* later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention
- \*X\* document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone
- \*Y\* document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art.
- \*G\* document member of the same patent family

Date of the actual completion of the international search

3 Apr11 2003

Date of mailing of the international search report

17/04/2003

Name and mailing address of the ISA

European Patent Office, P.B. 5818 Patentlaan 2  
NL - 2280 HV Rijswijk  
Tel. (+31-70) 340-2040, Tx. 31 651 epo nl,  
Fax: (+31-70) 340-3016

Authorized officer

Mazet, J-F

## INTERNATIONAL SEARCH REPORT

Int. Patent Application No.

PCT/IB 03/00074

## C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	WO 91 00298 A (FIREXTRA OY) 10 January 1991 (1991-01-10) abstract examples claims ----	1-7
X	WO 01 87988 A (THE PROCTER / GAMBLE COMPANY) 22 November 2001 (2001-11-22) abstract examples claims ----	1-7
A	WO 01 32751 A (COGNIS DEUTSCHLAND GMBH) 10 May 2001 (2001-05-10) claims; examples ----	1-7
A	PATENT ABSTRACTS OF JAPAN vol. 012, no. 186 (C-500), 31 May 1988 (1988-05-31) & JP 62 292802 A (HIGETA SHOYU KK), 19 December 1987 (1987-12-19) abstract & DATABASE WPI Section Ch, Week 198805 Derwent Publications Ltd., London, GB; Class D17, AN 1988-033032 & JP 62 292802 A (HIGETA SHOKYU KK), 19 December 1987 (1987-12-19) abstract ----	1-7
A	US 5 624 679 A (VOURNAKIS ET AL.) 29 April 1997 (1997-04-29) cited in the application column 15, line 4 - line 27 ----	1-7
A	DATABASE WPI Section Ch, Week 199816 Derwent Publications Ltd., London, GB; Class A11, AN 1998-177565 XP002235972 & RU 2 087 483 C (SOVA V V), 20 August 1997 (1997-08-20) abstract -----	

# INTERNATIONAL SEARCH REPORT

Int. Patent Application No  
PCT/IB 03/00074

Patent document cited in search report		Publication date	Patent family member(s)	Publication date
JP 63017901	A	25-01-1988	NONE	
WO 9100298	A	10-01-1991	FI 893223 A WO 9100298 A1 NO 910707 A RU 2046800 C1	31-12-1990 10-01-1991 22-02-1991 27-10-1995
WO 0187988	A	22-11-2001	PL 340132 A1 AU 6304601 A EP 1280829 A1 WO 0187988 A1	19-11-2001 26-11-2001 05-02-2003 22-11-2001
WO 0132751	A	10-05-2001	DE 19952073 A1 WO 0132751 A1 EP 1237988 A1	12-07-2001 10-05-2001 11-09-2002
JP 62292802	A	19-12-1987	JP 1033482 B JP 1549533 C	13-07-1989 09-03-1990
US 5624679	A	29-04-1997	US 5623064 A US 5622834 A AU 5917896 A WO 9639122 A1 US 2002106792 A1 US 2002098579 A1 US 2002091101 A1 US 6063911 A US 5846952 A US 5686115 A US 5858350 A US 5635493 A US 2001055807 A1 AU 695850 B2 AU 1296995 A CA 2177823 A1 CN 1142833 A EP 0731812 A1 JP 9506126 T NZ 277662 A TW 458987 B WO 9515343 A1	22-04-1997 22-04-1997 24-12-1996 12-12-1996 08-08-2002 25-07-2002 11-07-2002 16-05-2000 08-12-1998 11-11-1997 12-01-1999 03-06-1997 27-12-2001 27-08-1998 19-06-1995 08-06-1995 12-02-1997 18-09-1996 17-06-1997 27-04-1998 11-10-2001 08-06-1995
RU 2087483	C	20-08-1997	RU 2087483 C1	20-08-1997